# **AUTISM & ALUMINUM ADJUVANTS IN VACCINES**

How Aluminum Adjuvants in Vaccines Can Cause Autism



Published: August 18, 2017 (Version 1.0)

The Centers for Disease Control (CDC) asserts that vaccines and vaccine ingredients have been disproven as potential causes of autism. Statements by the CDC are generic and encompass all vaccines and vaccine ingredients. For example, the CDC states:

"Vaccines Do Not Cause Autism" "There is no link between vaccines and autism." "...no links have been found between any vaccine ingredients and autism spectrum disorder." (CDC website, August 2017)

These statements are not supported by available science. The CDC's evidence supporting these statements is limited to the MMR vaccine (Taylor 2014), thimerosal preservative (Taylor 2014) and vaccine antigen exposure (DeStefano 2013).

Dr Frank DeStefano of the CDC's Immunization Safety Office is co-author of a paper (Glanz 2015) which states:

"To date, there have been no populationbased studies specifically designed to evaluate associations between clinically meaningful outcomes and non-antigen ingredients, other than thimerosal."

This statement applies to, among other vaccine ingredients, aluminum adjuvant. Studies of MMR vaccine cannot be used as evidence of safety for other vaccines, for example vaccines that contain aluminum adjuvant. The overly-broad, generic assertions that no vaccines and no ingredients cause autism are thus not supported by scientific evidence. In fact, the CDC statements are contradicted by a large, consistent and growing body of scientific evidence, including:

1) studies showing neurotoxic and neuroinflammatory effects (e.g. microglial activation) from dosages of aluminum adjuvants lower than or approximately equal to dosages received by infants according to the CDC vaccine schedule (Crepeaux 2017, Petrik 2007, Shaw 2013, Shaw 2009);

2) studies linking vaccines to immune activation brain injury (Zerbo 2016, Li 2015);

3) studies showing that early-life immune activation is a causal factor in autism and other neurodevelopmental disorders and mental illnesses (e.g. schizophrenia) (Meyer 2009, Deverman 2009, Estes 2016, Kneusel 2014, Careaga 2017, Meyer 2014).

The accumulating evidence indicates that vaccine-induced immune activation, and aluminum adjuvants in particular, may cause mental illnesses and neurodevelopmental disorders, including autism. In this paper, we present scientific evidence that aluminum adjuvants can cause autism and other brain injuries. Also, we explain why the studies allegedly supporting the safety of aluminum adjuvants do not show safety for adverse neurological outcomes.

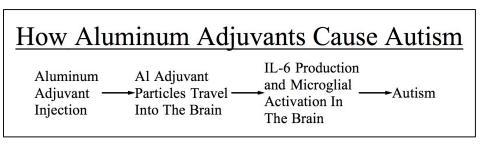


Fig 1: Proposed mechanism for how aluminum adjuvants cause autism. Each step is supported by replicated scientific studies.

## Immune Activation: A Cause of Autism and Mental Illness

The term "immune activation" describes the activation of the cellular components of the immune system. The developing brain can be injured by immune activation, with life-long consequences (Meyer 2009, Deverman 2009, Estes 2016, Kneusel 2014, Careaga 2017, Meyer 2014). Immune activation injury is linked to autism, schizophrenia, depression and other mental illnesses or neurodevelopmental disorders. Immune activation effects on the brain are mediated by immune system signaling molecules, especially cytokines (Estes 2016, Meyer 2014, Smith 2007, Choi 2016, Pineda 2013).

It is generally accepted that immune activation (e.g., from infection) during pregnancy is a risk factor for autism and schizophrenia in the offspring (Ciaranello 1995, Atladottir 2010, Brown 2012). The intensity and duration of immune activation and cytokine expression appear to be important factors influencing autism risk (Meyer 2014). Intense immune activation is associated with greater risk of autism (Careaga 2017, Atladottir 2010). Chronic inflammation is associated with greater risk of autism (Jones 2016, Zerbo 2014). However, there is no evidence that short-duration, lowintensity immune activation resulting from common childhood illnesses increase autism risk. Timing of immune activation in relation to stages of brain development is also an important factor (Meyer 2006, Meyer 2009).

Animal experiments have tested the effects immune activation of during pregnancy and postnatally on the development of offspring (Meyer 2009, Deverman 2009, Estes 2016, Kneusel 2014, Careaga 2017, Meyer 2014). In these experiments, pregnant animals (mice, rats and monkeys) or neonates are injected with a non-infectious immune activating substance such as "poly-IC" (which mimics a viral infection) or lipopolysaccharide (LPS, which mimics а bacterial infection). These substances cause immune system activation without infection. They induce fever and cytokine production and can have substantial effects on brain development if activation is sufficiently intense or prolonged and if exposure occurs during vulnerable developmental stages.

Immune activation has been demonstrated in mice to cause the three core behavioral symptoms of autism: decreased socialization and communication, and increased repetitive behaviors (Malkova 2012). Immune activation has also been shown to cause neuropathology (Weir 2015) and behavioral abnormalities in monkeys that resemble behaviors in human schizophrenia and autism (Bauman 2014, Machado 2015). See Fig. 2.

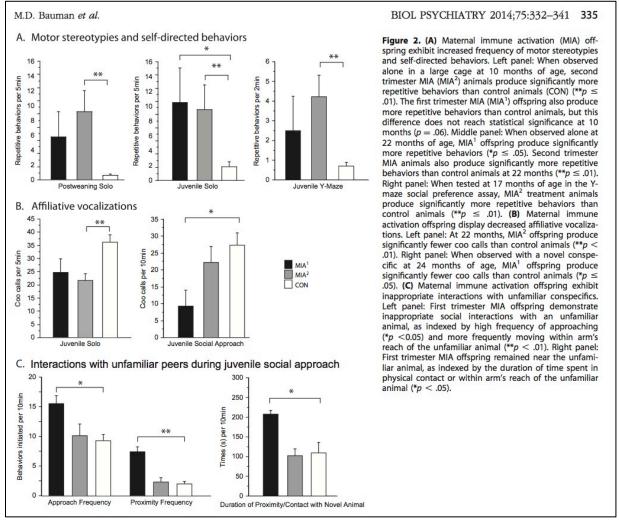


Fig 2: Maternal immune activation in monkeys caused behavioral abnormalities in juvenile offspring resembling behaviors in both autism and schizophrenia. MIA1 (Black)= first trimester immune activation; MIA2 (grey) 2nd trimester immune activation; CON (white) saline control. From Bauman et al. 2014

Immune activation also causes nonbehavioral effects associated with human autism (citations here link immune activation with these effects):

1) reduction in Purkinje cells (Shi 2009);

2) mitochondrial dysfunction (Giulivi 2013);

3) increase in brain volume (from IL-6 exposure, Wei 2012(b)) and neuron density in the brain (Smith 2012);

4) long term chronic brain inflammation (Garay 2012); and

5) microbiome disruption (dysbiosis) (Hsiao 2013).

These non-behavioral similarities further support the relevance of the immune activation models to human autism. The nonbehavioral (e.g., physiological) effects of immune activation have been reviewed (Labouesse 2015).

The cytokines interleukin-6 (IL-6) and interleukin-17a (IL-17) have been identified as mediating the behavioral effects of immune activation (Smith 2007, Malkova 2012, Choi 2016, Pineda 2013, Wei 2012(a), Wei 2013, Parker-Athill 2010, Wei 2016). The IL-6 findings have been replicated by different researchers using a variety of experimental methods. For example, in an experiment with poly-IC, abnormal behavior almost completely prevented is by simultaneous administration of IL-6-blocking antibody (Smith 2007, Pineda 2013). Injection of IL-6 by itself causes abnormal behavior that closely matches behavior resulting from poly-IC immune activation (Smith 2007). Inhibition of IL-6 signaling in a genetic autism model (BTBR mice) normalized social and repetitive behavior (Wei 2016). These results demonstrate that IL-6 is responsible for causing abnormal autism-like behavior.

The Patterson laboratory at CalTech was the first to report that IL-6 is responsible for causing the autism-like behavioral effects of immune activation (Smith 2007). Two papers from this research group state: "IL-6 is central to the process by which maternal immune activation causes longterm behavioral alterations in the offspring." (Smith 2007)

"...blocking IL-6 prevents >90% of the changes seen in offspring of poly(I:C)injected females, showing that gene expression changes, as well as behavioral changes, are normalized by eliminating IL-6 from the maternal immune response." (Smith 2007)

"IL-6 is necessary and sufficient to mediate these effects since the effects...are prevented by injection of pregnant mice with poly-IC combined with an anti-IL-6 antibody, and are mimicked by a single maternal injection of IL-6." (Garay 2013)

Brain exposure to elevated IL-6 by engineered virus showed that IL-6 exposure, initiated after birth, caused autism-like behaviors (Wei 2012(a)). The Wei 2012(a) paper states:

"We demonstrated that IL-6 is an important mediator of autism-like behaviors. Mice with an elevated IL-6 in brain developed autism-like behaviors, including impaired cognition ability, deficits in learning, abnormal anxiety-like trait and habituation, as well as a decreased social interaction initiated at later stages. These findings suggest that an IL-6 elevation in the brain could modulate certain pathological alterations and contribute to the development of autism." (Wei 2012(a))

More recent evidence shows that IL-17 acts downstream of IL-6 to cause autismlike behavioral abnormalities and atypical cortical development in mice (Choi 2016). Blocking either IL-6 or IL-17 prevents the autism-like behavior; an injection of IL-17 by itself causes the autism-like behavior (Choi 2016). IL-6 is known to induce IL-17 by promoting the development of Th17 cells which produce IL-17.

Immune activation animal models appear to be valid models for human neurological/psychiatric disorders, including autism (Estes 2016, Careaga 2017, Meyer 2014). The Estes 2016 review argues for the validity of the immune activation models to humans:

"These MIA (maternal immune activation) animal models meet all of the criteria required for validity for a disease model: They mimic a known diseaserelated risk factor (construct validity), they exhibit a wide range of disease-related symptoms (face validity), and they can be used to predict the efficacy of treatments (predictive validity)." (Estes 2016)

Evidence suggests a mediating role for IL-6 and IL-17 in human autism. For example, IL-6 is significantly elevated in the cerebellum in human autism (Wei 2011) and is highly elevated in some brain regions of some autistic individuals (Vargas 2005). Treatment of human autistics with the antiinflammatory flavonoid luteolin improves autistic behaviors in the individuals that also experience a decline in IL-6 blood levels (Tsilioni 2015). This result is consistent with a causal role for IL-6 in human autism. Also, IL-17 is elevated in human autism (Akintunde 2015, Al-Avadhi 2012, Suzuki 2011). Vitamin D reduces IL-17 production (Bruce 2011, Wobke 2014, Drozdenko 2014) and improves autistic behaviors in humans (Saad 2016, Jia 2015). The vitamin D findings

are consistent with a causal role for IL-17 in human autism.

IL-6 functioning appears to be similar or identical in mice and humans. No mousehuman differences in IL-6 functioning are described in a 2004 review (Mestas 2004). IL-6 functioning is quite conserved across species (Brown 2014). Central nervous system development in rodents and humans is governed by the same principles (Brown 2014). Hence, the fact that IL-6 causes autismlike behavioral abnormalities in animal models deserves a presumption of validity to humans.

Immune activation is a risk factor for autism, schizophrenia and other neurological/psychiatric disorders. The cytokines IL-6 and IL-17 are responsible for mediating the autism-like behavioral effects of immune activation in the animal models. The available evidence supports a causal role for IL-6 and IL-17 in human autism.

### Maternal vs. Postnatal Immune Activation

The timing of immune activation is an important factor influencing effects on the brain. The developing brain is vulnerable to immune activation injury; the mature, adult brain is apparently not nearly as vulnerable. Sensitivity to immune activation likely declines as the brain matures (Meyer 2014, Meyer 2007).

In most immune activation experiments, the offspring are exposed to immune activation during gestation (by stimulating the <u>maternal</u> immune system). In contrast, most vaccines are administered postnatally. This raises the question of whether <u>postnatal</u> immune activation can have similar effects on the brain as maternal immune activation. Diverse evidence indicates that the brain can be adversely affected by postnatal immune activation. Postnatal immune activation experiments, human case reports, and consideration of brain development timelines suggest that the human brain is vulnerable to immune activation injury for years after birth.

In the maternal immune activation experiments, inflammatory signaling and some cytokines (e.g. IL-6) traverse the placenta into the fetus. Consequently, immune activation in the mother causes immune activation and elevated cytokines in the fetus, and in the fetal brain (Oskvig 2012, Ghiani 2011).

Postnatal immune activation can have adverse neurological effects, including increased seizure susceptibility (Chen 2013, Galic 2008), learning and memory deficits (Harre 2008), and an increase in excitatory synapse formation (Shen 2016). Seizure disorders, learning and memory dysfunction, and elevated excitatory signaling are associated with autism.

Elevated IL-6 in the brain in the postnatal period causes neuronal circuitry imbalance mediates autism-like and behaviors in mice (Wei 2012(a)). The circuitry imbalance observed in Wei 2012(a) was an excess of excitatory synapses and a deficit of inhibitory synapses. See Fig. 3. Excessive excitatory signaling is observed in human autism (Robertson 2016, Freyberg 2015). In fact, an imbalance between excitatory and inhibitory signaling (towards excess excitation) has been posited as a central characteristic of autism (Robertson 2016, Freyberg 2015).

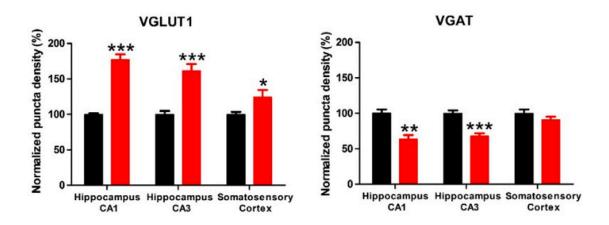


Fig 3: Elevation of IL-6 in the brains of mice (initiated shortly after birth) caused an increase in excitatory synapses (VGLUT1) and a decrease in inhibitory synapses (VGAT). Excessive excitatory signaling is observed in human autism. Red=Elevated IL-6; Black=Control. VGLUT1=excitatory synapses; VGAT=inhibitory synapses. \*P<0.05, \*\*P<0.01 and \*\*\*P <0.001. Adapted from Wei et al 2012(a).

In a maternal immune activation experiment with mice (Coiro 2015), autism-

relevant behavior and dendritic spine abnormalities (relevant to autism and schizophrenia) were ameliorated by administering an anti-inflammatory drug postnatally. The drug was started at birth and continued for 2 weeks, which roughly corresponds to age 2 in humans (Semple 2013). This result indicates that brain development is affected by postnatal inflammation, at times corresponding to when vaccines are given to humans.

Several case reports describe previously-healthy children that displayed sudden-onset autistic behavior during or subsequent to infection in the brain. All the cases had signs of intense brain inflammation. Here are brief descriptions:

*Delong 1981*: describes 3 children, ages 5, 7 and 11 with full-blown autistic behavior associated with brain inflammation. Brain inflammation was presumed in two cases and confirmed in one. The 5 and 7 year olds recovered completely, and the 11-year recovered partially.

*Marques* 2014: describes a previously healthy 32-month-old girl that suffered autistic regression from a viral central nervous system infection with associated brain inflammation.

*Ghaziuddin* 2002: describes a previously healthy 11-year-old boy that suffered permanent autistic regression after sudden onset herpes brain infection with associated brain inflammation.

*Gillberg* 1986: describes a previously healthy 14-year-old girl with permanent autistic regression from herpes brain infection with associated brain inflammation.

The most parsimonious explanation for these cases is that autistic behavior resulted from intense inflammation and cytokine production in the brain. Accordingly, these cases indicate that the human brain remains vulnerable to immune activation injury well into childhood, though the vulnerability almost certainly decreases with maturation. The susceptibility of older children to inflammation-induced autistic behavior strongly suggests that younger infants, of 0-2 years of age, are also vulnerable. It is not reasonable to claim, and there is no evidence to suggest, that the age range of 0-2 years (when most vaccines are given) is uniquely resistant to immune activation injury. All the available evidence indicates the opposite.

The immune activation experiments and case reports are consistent and indicate that immune activation and elevated cytokines in the postnatal period can cause brain injury.

The next critical question to consider is whether vaccines can cause immune activation and elevated cytokines in the brain.

### Postnatal Vaccination Affects Brain Development in Animal Model

The first study to test the effect of postnatal vaccination on brain development was published in 2015 (Li 2015). In this experiment, neonatal rats were administered bacillus calmette-guerin (BCG) vaccine, hepatitis B (HBV) vaccine or a combination (BCG+HBV) timed to imitate human infant vaccination schedules. BCG and HBV vaccines produced opposite effects on the brain. Specifically, BCG enhanced synaptic plasticity and long-term potentiation (LTP, the basis for learning and memory); HBV inhibited synaptic plasticity and LTP. BCG and HBV vaccines also caused opposite changes in some synapse protein levels.

HBV vaccine (but not BCG vaccine) increased IL-6 gene expression in the brain;

increased gene expression likely indicates an elevation in brain IL-6. The HBV vaccine contains aluminum adjuvant, and the BCG does not contain aluminum adjuvant. Hence, the aluminum adjuvant may be the ingredient responsible for the elevated IL-6 gene expression. See Fig. 4.

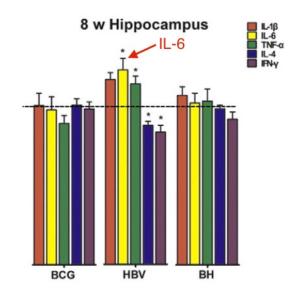


Fig. 4: Hepatitis B vaccine, but not BCG vaccine, increased IL-6 gene expression in the brain at 8 weeks after neonatal vaccination. Hepatitis B vaccine contains aluminum adjuvant; BCG vaccine does not. Elevated IL-6 causes autism-like behaviors in animal models. \*P<0.05 Adapted from Li et al 2015.

The Li et al study showed that the vaccines caused other changes in the brain, including 1) changes in long-term potentiation (LTP) (Hep B decreased LTP), 2) changes in dendritic spines, and 3) changes in synapse protein expression. Changes in synapse proteins and dendritic spines have been observed in human brain disorders.

Li et al. attribute the brain effects to changes in cytokine levels and immune polarization (Th1/Th2 polarization) induced by the vaccines. Aluminum adjuvants cause Th2 polarization. Li et al. state that the results suggest vaccines can interact by way of immune activation effects:

"...our data suggested that combinations of different vaccines can mutually interact (enhance or counteract). The mechanism of synaptic plasticity modulation through neonatal BCG/HBV vaccination may be via systemic Th1/Th2 bias accompanied by a specific profile of cytokines and neurotrophins in the brain." (Li 2015)

Li 2015 demonstrates that vaccines affect brain development by an immune activation mechanism. Further, since aluminum adjuvants induce Th2 activation and long term Th2 polarization, the Li 2015 results suggest that all aluminum-adjuvanted vaccines may cause adverse effects similar to the HBV vaccine. Accordingly, the Li 2015 results suggest that studies showing that immune activation causes neurological/psychiatric disorders are relevant to vaccine adverse effects.

## Vaccines Are Given During Synaptogenesis

Another way to answer the question of brain vulnerability to immune activation is to consider the types of brain development processes occurring when vaccines are administered. Vaccines are given primarily in the first 18 months after birth. The human undergoes brain intense and rapid development this period. during Synaptogenesis (formation synapse of connections between neurons) is especially intense in this period.

The vulnerability of the developing brain to immune activation is apparently related to the specific types of brain development processes occurring (Tau 2010, Meyer 2006, Meyer 2007). Such processes include migration (movement of neurons to final locations in the brain), adhesion (formation of chemical-mechanical attachments between brain cells), and synaptogenesis (formation of synapse connections between neurons), among others (neurogenesis, gliogenesis, myelination etc).

Cytokines affect brain development processes. For example, elevated IL-6 affects migration, adhesion and synaptogenesis (Wei 2011). Elevated IL-6 in the postnatal period promotes an excess of excitatory synapses and a deficit of inhibitory synapses, and mediates autism-like behaviors (Wei 2012(a)).

In humans, a dramatic increase in synaptogenesis begins around the time of birth, and continues until about age 3 (Huttenlocher 1997, Tau 2010, Stiles 2010, Semple 2013). Vaccines are administered during this intense synaptogenesis. See Figs. 5-6. Elevated brain IL-6 induced by vaccination during synaptogenesis may cause an excitatory-inhibitory imbalance, towards excitation. An excitatory imbalance has been observed in human autism (Robertson 2016, Freyberg 2015).

Synaptogenesis tapers off through childhood and adolescence. This fact may explain why some older children and teens can suffer autistic regression after intense brain inflammation, but apparently become less vulnerable to immune activation brain injury with age.

Intense synaptogenesis occurs at ages 0-18 months, when many vaccines are administered. Consequently, vaccines may adversely impact synaptogenesis if they induce inflammation or IL-6 in the brain.

The timing of brain development processes in humans supports the idea that the human brain is vulnerable to immune activation and cytokines in the first few years after birth, when vaccines are administered. Disruption of synaptogenesis by vaccineinduced immune activation is a particular concern.

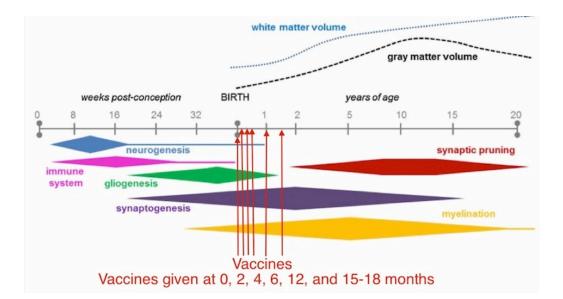
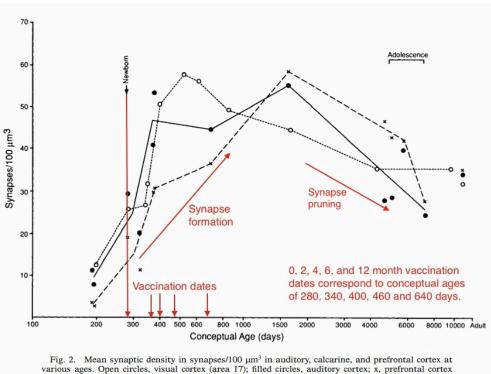


Fig. 5: Timeline of specific brain developmental processes in humans. Synaptogenesis is most intense during the first couple years of life, when vaccines are administered. Timing of vaccination according to the CDC vaccine schedule is shown. Elevated IL-6 during synaptogenesis may cause an excitatory-inhibitory synapse imbalance, towards excitation. Adapted from Semple 2013.



(middle frontal gyrus).

Fig. 6: Measurements of synapse density in human cadavers of various ages indicate a dramatic increase in synapses in the first few years of life. Vaccines are administered during intense synapse formation. Elevated IL-6 during synaptogenesis may cause an excitatory-inhibitory synapse imbalance, towards excitation. Image adapted from Huttenlocher and Dabholkar 1997.

## Aluminum Adjuvants: Neurotoxic At Vaccine Dosages

Aluminum (Al) adjuvants have an essential role in many vaccines: to stimulate immune activation. Without Al adjuvants, these vaccines would have greatly reduced efficacy.

Aluminum adjuvants comprise submicron particles (primary particles) of aluminum compounds, typically AlOH, AlPO4, AlSO4 or mixtures. The primary particles are typically agglomerated into larger particles with sizes of about 2-20 microns (Harris 2012). The Al adjuvant materials have low solubility in water and body fluids. Al adjuvant particles are biopersistent and can remain in the body for months or years (Flarend 1997, Khan 2013, Gherardi 2001).

Aluminum ingested in the diet has low oral absorption (about 0.3%), is rapidly excreted by the kidneys, is (mostly) excluded from the brain by the blood-brain barrier, and is in a solubilized, Al3+ ionic form (not particulate) These defenses are adequate for protecting the brain from natural levels of exposure. aluminum These protective mechanisms are unable to protect the brain from injected aluminum adjuvant particles. Al adjuvant particles are too large to be removed by the kidneys, and are carried blood-brain across the barrier by macrophages.

Dosages of aluminum adjuvants received by infants according to the CDC vaccination schedule are:

#### Birth (Hep B):

74 mcg/kg (250 mcg for 3.4 kg infant)

#### 2 month:

245 mcg/kg (1225 mcg for 5 kg infant)

#### 4 month:

150 mcg/kg (975 mcg for 6.5 kg infant)

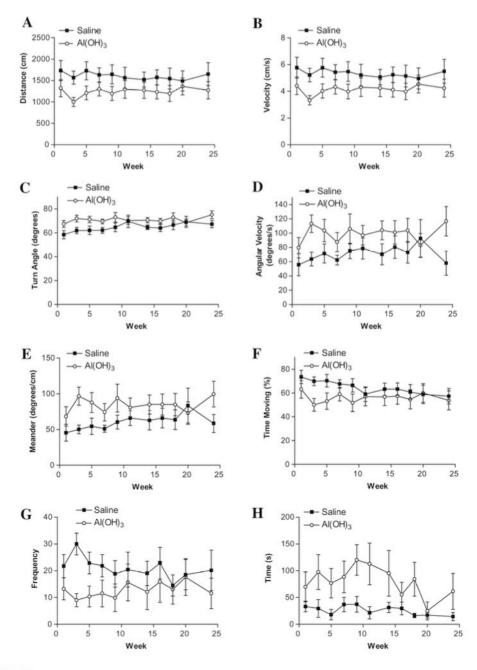
#### 6 month:

153 mcg/kg (1225 mcg for 8 kg infant)

These are maximum-possible dosages (because different vaccine products have different amounts) for average-weight infants.

Accumulating evidence shows that aluminum adjuvants have adverse neurological effects at dosages lower than or approximately equal to dosages infants receive from vaccines. These effects appear to depend on the particulate nature and biopersistence of the aluminum adjuvant. Injected Al adjuvant has adverse effects that are apparently mediated by the particles and independent of solubilized Al3+ ions released by the slowly dissolving particles (Crepeaux 2017).

Al adjuvant injections in mice cause adverse effects at vaccine-relevant dosages of 100, 200, 300 and 550 mcg/Kg body weight (Crepeaux 2017, Shaw 2009, Petrik 2007, Shaw 2013). These include deficits in learning and memory (Shaw 2009), deficits in neuromuscular strength/function (Petrik 2007), and changes in locomotor activity and/or gait (Shaw 2009, Shaw 2013). Autism is associated with gait and movement abnormalities (Kindregan 2015) and memory dysfunction (Williams 2006).



#### Fig. 4.

Open field movement analysis as an assessment of spontaneous activity and anxiety in control mice vs. mice injected six times with aluminum hydroxide. Aluminum hydroxide injected mice showed the following behavioural changes: (A) Shorter distances moved (\*\*\*p < 0.0001). (B) Slower movement (\*\*\*p < 0.0001). (C) Greater mean turn angle (\*\*\*p < 0.0001). (D) More rapid turning (\*\*\*p < 0.0001). (E) Greater meander (\*\*\*p < 0.0001). (F) Smaller percentage of time in overall movement (\*\*p = 0.0030). (G) Fewer entries into the centre of the open field (\*\*\*p < 0.001). Late entry into centre (\*\*\*p < 0.0001). (All measures, two-way ANOVA).

Fig. 7: Dosage of 300mcg/Kg AlOH adjuvant caused large and persistent changes in exploratory behavior and movement in open field tests. This is an indicator of neurotoxicity. Human autistics also display abnormal movement and exploratory behavior. Adapted from Shaw and Petrik 2009.

Al adjuvant dosages of 200mcg/Kg (as 3 x 66mcg/Kg) (Crepeaux 2017) and 300mcg/Kg (as 6 x 50mcg/Kg) (Shaw 2009) increased microglial activation in the ventral forebrain and lumbar spinal cord, respectively. The elevated microglial activation was measured about 6 months after Al adjuvant injection, which suggests that the microglial activation is chronic. Activated microglia indicate an ongoing inflammatory process and suggest the presence of elevated cytokines. Human autistics have activated microglia and cytokines throughout the brain elevated Suzuki 2013, (Vargas 2005, 2009). Li

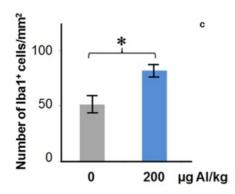


Fig. 8: Al adjuvant (200mcg/Kg) caused an increase in microglial activation in the brain of mice. The protein iba1 indicates activated microglia. Measurements were performed 6 months after Al adjuvant injection, indicating that the microglial activation is a chronic condition. \* P<0.05. From Crepeaux et al., 2017.

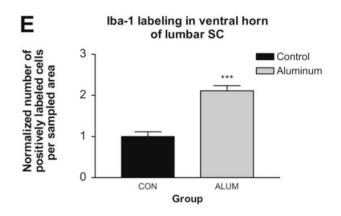


Fig. 9: Al adjuvant (300mcg/Kg) caused an increase in microglial activation in the lumbar spinal cord of mice. The protein iba1 indicates activated microglia. Measurements were performed 6 months after Al adjuvant injection, indicating that the microglial activation is a chronic condition. \*\*\*p < 0.001, one-way ANOVA. From Shaw and Petrik 2009.

Activated microglia are implicated as a causal factor in autism, because microglia mediate inflammation in the brain. Microglia can produce IL-6 when in an activated state. A recent review on microglia and autism (Takano 2015) states:

"...any factors that alter the number or activation state of microglia either in utero or during the early postnatal period can profoundly affect neural development, thus resulting in neurodevelopmental disorders, including autism." (Takano 2015)

Microglia appear to play an important role in the causation of autism (Takano 2015, Kneusel 2014). Hence, the microglial activation caused by aluminum adjuvants suggests a role in autism.

Several studies show that Al adjuvants increase brain aluminum content (Crepeaux 2017, Flarend 1997, Shaw 2009, Khan 2013, Crepeaux 2015). A dosage of 200 mcg/Kg Al adjuvant caused a 50-fold increase in brain aluminum content in mice, from 0.02 ug/g to 1.0 ug/g dry weight of brain (Crepeaux 2017). These measurements were

performed 6 months after the final injection, indicating that the Al persists in the brain long-term (Crepeaux 2017). See Fig. 10. Al adjuvants have been found to accumulate in the brain of mice up to one year after injection (Khan 2013). Crepeaux 2015 demonstrated persistence and increasing accumulation of Al adjuvant particles up to 270 days in spleen and lymph nodes of mice. Increasing accumulation of Al in distant organs over time suggests that toxic effects may increase with time, and may be delayed by months or years after exposure.

The 400 and 800 mcg/Kg doses used in the Crepeaux 2017 study did not cause adverse effects or elevated brain aluminum. The authors attribute this surprising inverted dose-response relationship to granulomas induced by the higher dosages. Granulomas trap the Al adjuvant at the injection site, thereby preventing its transport into the brain and other sensitive tissues. Granulomas occur after about 1% of vaccinations (Bergfors 2014). This is cause for concern because it indicates that, for 99% of vaccinations, the Al adjuvant can be transported around the body. It is not confined to a granuloma. See Fig. 11.

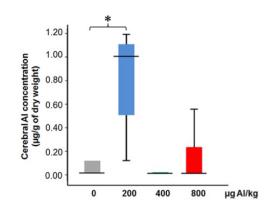


Fig. 10: Dosage of 200 mcg/Kg Al adjuvant caused a 50-fold increase in brain aluminum content, from 0.02 to 1.00 ug/g dry weight, in mice. Higher dosages (400 and 800 mcg/Kg) did not increase brain Al

content, presumably because the higher dosages caused a granuloma at the injection site. A granuloma traps the Al adjuvant at the injection site, thereby preventing systemic dispersal and transport into the brain. These measurements were performed 6 months after the final injection, indicating that the Al persists in the brain long-term. \*P<0.05. From Crepeaux et al., 2017.

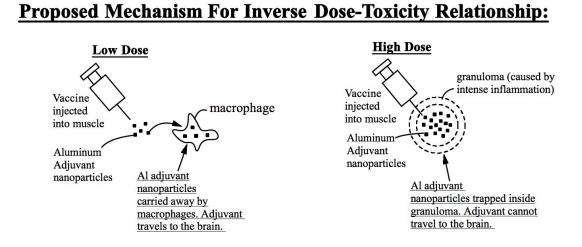


Fig. 11: High dose Al adjuvant injection into the muscle causes a granuloma, which traps the Al adjuvant and prevents it from traveling into the brain. Low dose does not form a granuloma. Hence, the lower dose is free to travel to the brain. Consequently, the lower dose is more toxic than the higher dose. This mechanism explains the surprising inverted dose-toxicity results of Crepeaux et al. 2017.

### Particle Transport and Macrophage Chemotactic Protein (MCP-1)

Aluminum adjuvants travel into the brain (Khan 2013, Crepeaux 2015, Crepeaux 2017, Shaw 2009, Flarend 1997). Al adjuvant particles are carried through the blood-brain barrier and into the brain by macrophages (Khan 2013). Transport is promoted by macrophage chemotactic protein-1 (MCP-1) (Khan 2013). MCP-1 causes macrophages to travel around the body and into the brain. Particle transport into the brain by macrophages is well-established and has been investigated for therapeutic applications (Choi 2012, Pang 2016).

MCP-1 is elevated in the brains of human autistics (Vargas 2005) and is elevated in the blood of neonates later diagnosed with autism (Zerbo 2014). This suggests that neonates with high MCP-1 will experience elevated Al adjuvant transport into the brain when injected with Al adjuvanted vaccines. This is consistent with Al adjuvants causing autism by inducing immune activation and elevated cytokines in the brain.

### Aluminum Induces IL-6 Expression In The Brain

Water-soluble aluminum salts (e.g. AlCl3, Al lactate) induce elevated IL-6 in the

brain and other tissues. In fact, aluminum appears to selectively induce IL-6 (Viezeliene 2013). Studies of aluminum exposure and IL-6 expression in the brain include:

Cao 2016: Ingestion of 30 or 90 mg/kg/day aluminum (as AlCl3) for 90 days significantly increased gene expression of IL-6 and other cytokines in the brain (hippocampus).

Alawdi 2016: Ingestion of 3.4 mg/kg/day aluminum (as AlCl3) for 6 weeks caused a 4-fold increase in IL-6 in the brain (hippocampus). This dosage is far lower than the outdated "no observed adverse effects level" (NOAEL) oral dosages (26 and 62 mg/kg/day) used as benchmarks for toxicity threshold (Mitkus 2011, Offit 2003).

In fact, other experiments show that oral dosages of 3.4, 4, 5.6, 6, and 20.2 mg/Kg/day aluminum cause numerous adverse effects in mice or rats and hence the NOAEL for orally ingested Al is currently unknown (Alawdi 2016, Dera 2016, Sethi 2008, Sethi 2009, Bilkei-Gorzo 1993).

The induction of IL-6 may occur because aluminum strongly induces oxidative stress (Exley 2003). Oxidative stress induces IL-6 expression (Viezeliene 2013).

## CDC Website Cites Fatally Flawed Study Of Al Adjuvants (Mitkus 2011)

Dosages of Al adjuvants received by infants increased dramatically as the vaccine schedule was expanded in the 1980s and 1990s. However, as the vaccine schedule expanded, the increasing dosages of Al adjuvants were not tested for safety. Government agencies (HHS, NIH, CDC, FDA) have not pursued any new experimental work on Al adjuvant toxicity.

To support the safety of Al adjuvants at today's higher dosages, the CDC cites a 2011 FDA study of aluminum exposure from vaccines (Mitkus 2011). This study is the only scientific evidence cited by the CDC and FDA websites to support the safety of Al adjuvants.

The Mitkus 2011 study is a theoretical modeling study of Al adjuvant kinetics; it contains no new data concerning Al adjuvant models toxicity (from animal or epidemiology). Mitkus 2011 calculates a body burden of aluminum resulting from the slow dissolution of Al adjuvant particles, and compares the dissolved-aluminum body burden to a "minimal risk level" (MRL). The MRL is derived from a study of ingested Al toxicity in mice (Golub 2001). The Golub 2001 study provides the NOAEL (26 mg/kg/day ingested), which is converted into the MRL for human infants (based on 1mg/kg/day ingested) by using a safety factor of about 30.

The Mitkus study is fatally flawed for these reasons:

### 1) MITKUS ASSUMES AL ADJUVANT PARTICLES ARE HARMLESS

Mitkus makes an unstated assumption that Al adjuvants have zero toxicity while in particulate form. Mitkus only considers the potential toxicity of aluminum ions (Al3+) released by the slowlydissolving Al adjuvant particles.

Al adjuvants comprise low-solubility and biologically-persistent microscopic particles. The Mitkus analysis assumes that the particles are absolutely nontoxic and perfectly harmless, even when present in the brain and other organs. Mitkus provides no justification for this unstated assumption. Further, the assumption is contradicted by recent findings on Al adjuvant toxicity (Crepeaux 2017) and particulate toxicity generally. Particles can have toxic effects mediated by surface chemistry (e.g. surface charge and surface catalytic activity) and particle shape, among other characteristics of solid particles (Sharifi 2012, Podila 2013).

Several studies show injected Al adjuvants cause behavioral abnormalities, abnormal weight gain, learning and memory impairment, motor neuron death/apoptosis, neuromuscular strength deficits, chronic microglial activation/brain inflammation, and large (e.g. 50X) increases in brain and spinal cord aluminum content (Petrik 2007, Shaw 2009, Shaw 2013, Crepeaux 2017). These adverse effects occur at dosages less than or approximately equal to dosages received by infants according to the CDC vaccine schedule.

### 2) NEW RESEARCH SHOWS INGESTED AL HARMFUL AT DOSAGES LOWER THAN 26 MG/KG/DAY

Mitkus assumes that Al adjuvant toxicity is mediated exclusively by solubilized Al (Al3+ ions) released by the slowly-dissolving Al adjuvant particles. To establish a threshold toxicity level from the solubilized Al, Mitkus relies on a mouse feeding study (Golub 2001) reporting a "noobserved adverse effects level" (NOAEL) oral dosage of 26 mg/Kg/day ingested aluminum. Mitkus used a 30X safety factor for applying this dosage to humans, which is reasonable.

However, other experiments show that much lower oral dosages of 3.4, 4, 5.6, 6, and 20.2 mg/Kg/day aluminum cause adverse effects in mice or rats (Alawdi 2016, Dera 2016, Sethi 2008, Sethi 2009, Bilkei-Gorzo 1993). The adverse effects include chronic brain inflammation, learning and memory impairment, and kidney inflammation. So, the Mitkus analysis is wrong because 26 mg/kg/day is not a NOAEL. The "minimal risk level" (MRL) determined by Mitkus is too high by a factor of at least 26/3.4 = 7.6. Using a corrected NOAEL of 3.4 mg/Kg/day (based on Alawdi 2016) results in vaccine aluminum exposure exceeding the MRL for AlPO4 adjuvant, and approximately matching the MRL for AlOH adjuvant. The new, corrected MRL lines indicate that Al phosphate adjuvant (Fig. 12) and Al hydroxide adjuvant (Fig. 13) from the CDC vaccine schedule may cause toxicity from the solubilized Al per se.

Since 3.4mg/Kg/day is not a NOAEL (adverse effects were observed at this dosage) the true NOAEL is less than 3.4/mg/Kg/day. See Figs. 12-13.

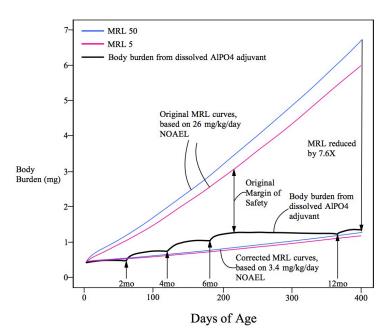


Fig. 12: Body burden vs. MRL comparison chart for Al <u>phosphate</u> adjuvant (AlPO4) corrected in accordance with the new discovery (Alawdi 2016) that ingestion of 3.4 mg/kg/day Al causes adverse effects. The body burden exceeds the corrected MRL curve for almost the entire first year of life, indicating toxicity. The toxicity of Al adjuvant <u>particles</u> is a separate, additional issue. MRL 50 and MRL 5 refer to two different infant growth rates. Adapted from Mitkus et al., 2011.

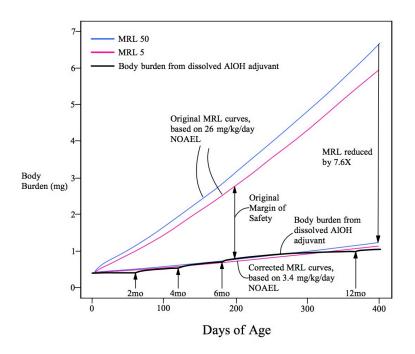


Fig. 13: Body burden vs. MRL comparison chart for Al <u>hydroxide</u> adjuvant (AlOH), corrected in accordance with the new discovery (Alawdi 2016) that ingestion of 3.4 mg/kg/day Al causes adverse effects. The body burden overlaps the new, corrected MRL, indicating borderline toxicity. The margin of safety is gone. MRL 50 and MRL 5 refer to two different infant growth rates. The toxicity of Al adjuvant <u>particles</u> is a separate, additional issue. Adapted from Mitkus et al., 2011.

### 3) NO AL ADJUVANT TOXICITY DATA CITED, DESPITE AVAILABILITY

Mitkus does not cite any toxicity data for injected Al adjuvants. Mitkus instead uses toxicity data for ingested, non-particulate, water-soluble Al (Golub 2001, which used Al lactate) to derive the MRL. This data comes from a single study (Golub 2001).

So, remarkably, Mitkus claims a safe level of injected Al adjuvant exposure, <u>without citing any Al adjuvant toxicity data</u>. The error is unnecessary and neglectful because at least two animal studies of injected Al adjuvant toxicity were available prior to the Mitkus publication in 2011 (Petrik 2007, Shaw 2009). These papers were not cited or mentioned by Mitkus 2011.

Each of these three flaws is fatal for the validity of the Mitkus study in establishing the safety of aluminum adjuvants. Hence, the CDC is completely lacking valid evidence for the safety of Al adjuvants. This is especially true for safety regarding neurological and long-term outcomes, because other available studies of Al adjuvant safety (e.g., Jefferson 2004) do not consider (or are incapable of detecting) these outcomes.

## CDC Fails To Investigate Toxicity of Al Adjuvants

The CDC has conducted no epidemiological studies on long term safety (e.g. considering neurological outcomes) of Al adjuvants. There is one ecological study of country-level data, which reported an association between Al adjuvant exposure and autism (Tomljenovic 2011). However, being an ecological study, it is highly susceptible to confounding and biases.

Dr Frank DeStefano of the CDC's Immunization Safety Office is co-author of a feasibility study (Glanz 2015) on using the Vaccine Safety Datalink (VSD) to investigate the safety of individual vaccine ingredients. The paper focuses on Al adjuvants. It acknowledges that thimerosal is the only vaccine ingredient studied for autism or neurological safety, and that a possible association between Al adjuvants and autism has not been explored in epidemiological studies. Glanz 2015 states:

"To date, there have been no population-based studies specifically designed to evaluate associations between clinically meaningful outcomes and nonantigen ingredients, other than thimerosal."

The CDC has not investigated Al adjuvant safety concerns, despite the accumulating scientific evidence of harm and evidence linking Al adjuvants to immune activation mechanisms of brain injury.<sup>1</sup>

<sup>&</sup>lt;sup>1</sup> However, the Glanz paper notes that studies of aluminum adjuvants are problematic because of expected small differences in

adjuvants, especially in relation to autism and other long term neurological outcomes.

### Conclusion

The science reviewed here tells a consistent and compelling story: that vaccines may cause autism by stimulating immune activation and elevated cytokines in the brain. Al adjuvants are implicated as a cause of autism because they can be transported into the brain, because they cause microglial activation at vaccine-relevant dosages, and because aluminum induces IL-6 in the brain.

In statements asserting no vaccineautism link, the CDC cites scientific evidence that is not relevant to Al adjuvant safety or is incapable of disproving an Al adjuvantautism link (Taylor 2014, DeStefano 2013, Mitkus 2011). In support of claims for Al adjuvant safety, the CDC relies on a profoundly flawed theoretical modelling study (Mitkus 2011). There is little scientific evidence supporting the safety of Al

exposures in the low and high exposure groups. Glanz 2015 concludes: "...children below the 10th percentile would be exposed to between 0 mg and 3.1mg, while children above the 90th percentile would be exposed to between 4.8 mg and 5.3 mg of aluminum from vaccines. It is unclear if such differences in aluminum exposure would be biologically meaningful." (Glanz 2015). So, epidemiological studies may not provide reliable evidence for safety or harm. Controlled, prospective human trials of aluminum adjuvant exposure from vaccines will likely be prohibited for ethical reasons. Also, Al adjuvants are essential ingredients for Al adjuvanted vaccines. Consequently, it will be challenging to design studies of long term adverse effects of Al adjuvants in humans. Experiments in animal models can provide valuable information. Al adjuvants should be tested for effects on: 1) excitatory/inhibitory imbalance; 2) core symptoms of autism (social, communicative and repetitive/stereotyped behaviors); 3) IL-6, IL-17, and other cytokine levels in the brain; 4) other physiological abnormalities associated with autism (e.g. mitochondrial dysfunction, microbiome dysbiosis, Purkinje cell loss, cerebellum abnormalities etc); and 5) microglial activation and immune activity in the brain. Investigating these outcomes can provide valuable information concerning the safety of Al adjuvants.

### References

Akintunde et al., 2015 Increased production of IL-17 in children with autism spectrum disorders and co-morbid asthma, Journal of Neuroimmunology 286 (2015) 33-41.

Al-Ayadhi et al., 2012 Elevated serum levels of interleukin-17A in children with autism, Journal of Neuroinflammation 2012, 9:158.

Alawdi et al., Neuroprotective Effect of Nanodiamond in Alzheimer's Disease Rat Model: a Pivotal Role for Modulating NF-κB and STAT3 Signaling, Molecular Neurobiology, 54 (3):1906-1918.

Atladottir et al., Maternal infection requiring hospitalization during pregnancy and autism spectrum disorders, Journal of Autism and Developmental Disorders, 2010 Dec;40(12):1423-1430.

Bauman et al., 2014 Activation of the Maternal Immune System During Pregnancy Alters Behavioral Development of Rhesus Monkey Offspring, Biological Psychiatry, 2014;75: 332–341

Bergfors et al., 2014 How common are long-lasting, intensely itching vaccination granulomas and contact allergy to aluminium induced by currently used pediatric vaccines? A prospective cohort study, European Journal of Pediatrics, 173:1297–1307.

Bilkei-Gorzo, 1993, Neurotoxic effect of enteral aluminum, Food and Chemical Toxicology, 31(5):357-361.

Brown et al., 2014 Metabolic consequences of interleukin-6 challenge in developing neurons and astroglia, Journal of Neuroinflammation, 11:183.

Brown et al., Epidemiologic studies of exposure to prenatal infection and risk of schizophrenia and autism, Developmental Neurobiology, 2012 October ; 72(10): 1272–1276.

Bruce et al., 2011 Converging pathways lead to overproduction of IL-17 in the absence of vitamin D signaling, 2011 Aug; 23(8): 519–528.

Careaga et al 2017 Maternal Immune Activation and Autism Spectrum Disorder: From Rodents to Nonhuman and Human Primates, Biological Psychiatry, March 1, 2017; 81:391–401.

Chen et al., Postnatal systemic inflammation exacerbates impairment of hippocampal synaptic plasticity in an animal seizure model, Neuroimmunomodulation, 2013;20(4):223-32.

Choi et al., 2012, Delivery of nanoparticles to brain metastases of breast cancer using a cellular Trojan horse, Cancer Nanotechnology, 3:47–54.

Choi et al., 2016 The maternal interleukin-17a pathway in mice promotes autismlike phenotypes in offspring, Science, 2016 Feb 26; 351(6276): 933–939.

Ciaranello et a The Neurobiology of Infantile Autism, The Neuroscientist, 1:361-367

Coiro et al., Impaired synaptic development in a maternal immune activation mouse model of neurodevelopmental disorders, Brain, Behavior, and Immunity, Nov;50:249-258.

Crepeaux et al., 2015 Highly delayed systemic translocation of aluminum-based adjuvant in CD1 mice following intramuscular injections, Journal of Inorganic Biochemistry, 152:199-205.

Crepeaux et al., 2017 Non-linear dose-response of aluminium hydroxide adjuvant particles: Selective low dose neurotoxicity, Toxicology, 375 (2017) 48–57.

DeLong et al., 1981 Acquired reversible autistic syndrome in acute encephalopathic illness in children, Archives of Neurology, 36:191-194.

Dera 2016, Protective effect of resveratrol against aluminum chloride induced nephrotoxicity in rats, Saudi Medical Journal, 37 (4).

DeStefano et al., 2013 Increasing Exposure to Antibody-Stimulating Proteins and Polysaccharides in Vaccines Is Not Associated with Risk of Autism, The Journal of Pediatrics, 163 (2).

Deverman and Patterson, 2009 Cytokines and CNS Development, Neuron 64:61-78.

Drozdenko et al., 2014 Oral vitamin D increases the frequencies of CD38+ human B cells and ameliorates IL-17-producing T cells, Experimental Dermatology, 23: 107-112.

Estes and McAllister, 2016 Maternal immune activation: implications for neuropsychiatric disorders, Science, 353 (6301) 772-777.

Exley, 2003 The Pro-Oxidant Activity of Aluminum, Free Radical Biology and Medicine, 36(3): 380-387.

Flarend et al., 1997 In vivo absorption of aluminum-containing vaccine adjuvants using 26 Al, Vaccine, 15(12/13):1314-1318.

Freyberg et al., 2015 Reduced perceptual exclusivity during object and grating rivalry in autism, Journal of Vision, 15(13):11, 1–12.

Galic et al., 2008 Postnatal Inflammation Increases Seizure Susceptibility in Adult Rats, The Journal of Neuroscience, 2008, 28 (27) 6904-6913.

Garay et al., 2013 Maternal immune activation causes age- and region-specific changes in brain cytokines in offspring throughout development, Brain, Behavior, and Immunity, 31: 54-68.

Ghaziuddin et al., 2002 Autistic symptoms following herpes encephalitis, European Child and Adolescent Psychiatry, Vol. 11, No. 3:142-146.

Gherardi et al., 2001 Macrophagic myofasciitis lesions assess long-term persistence of vaccine-derived aluminium hydroxide in muscle, Brain, 124:1821-1831.

Ghiani et al., 2011 Early effects of lipopolysaccharide induced inflammation on foetal brain development in rat, ASN Neuro, 3 (4): 233-245.

Gillberg 1986 Brief Report: Onset at Age 14 of a Typical Autistic Syndrome. A Case Report of a Girl with Herpes Simplex Encephalitis, Journal of Autism and Developmental Disorders, VoL 16, No. 3:369-375.

Giulivi et al 2013 Gestational Exposure to a Viral Mimetic Poly(I:C) Results in Long-Lasting Changes in Mitochondrial Function by Leucocytes in the Adult Offspring, Mediators of Inflammation, Vol 2013:609602.

Glanz et al., 2015, Cumulative and episodic vaccine aluminum exposure in a populationbased cohort of young children, Vaccine 33:6736–6744.

Golub et al., 2001 Long-term consequences of developmental exposure to aluminum in a suboptimal diet for growth and behavior of Swiss Webster mice, Neurotoxicology and Teratology 23 (2001) 365–372.

Gupta et al., 1998 Th1- and Th2-like cytokines in CD4+ and CD8+ T cells in autism, Journal of Neuroimmunology, 85:106-109.

Harre et al., 2008 Neonatal inflammation produces selective behavioural deficits and alters *N*-methyl-D-aspartate receptor subunit mRNA in the adult rat brain, European Journal of Neuroscience, 2008 Feb; 27(3): 644–653.

Harris et al., 2012 Alhydrogel® adjuvant, ultrasonic dispersion and protein binding: A TEM and analytical study, Micron, 43:192-200.

Hsiao et al., 2013 The microbiota modulates gut physiology and behavioral abnormalities associated with autism, Cell, 155(7): 1451-1463.

Huttenlocher and Dabholkar, 1997 Regional Differences in Synaptogenesis in Human Cerebral Cortex, Journal of Camparative Neurology, 387:167–178 (1997).

Jefferson 2004 Adverse events after immunisation with aluminium-containing DTP vaccines: systematic review of the evidence, The Lancet 4:84-90.

Jones et al., 2016 Autism with Intellectual Disability is Associated with Increased Levels of Maternal Cytokines and Chemokines During Gestation, Molecular Psychiatry, 22(2):273-279.

Khan et al., 2013 Slow CCL2-dependent translocation of biopersistent particles from muscle to brain, BMC Medicine, 11:99.

Kindregan et al., 2015 Gait Deviations in Children with Autism Spectrum Disorders: A Review, Autism Research and Treatment, ID:741480.

Knuesel et al., 2014, Maternal immune activation and abnormal brain development across CNS disorders, Nature Reviews 10:643-660.

Labouesse et al., 2015, Long-term pathological consequences of prenatal infection: beyond brain disorders, American Journal of Physiology, 309:1.

Li et al. 2009 Elevated Immune Response in the Brain of Autistic Patients, Journal of Neuroimmunology, 207(1-2): 111–116.

Li et al., 2015 Neonatal vaccination with bacillus Calmette–Guérin and hepatitis B vaccines modulates hippocampal synaptic plasticity in rats, Journal of Neuroimmunology, 288 (2015) 1-12.

Machado et al., 2015 Maternal Immune Activation in Nonhuman Primates Alters Social Attention in Juvenile Offspring, Biological Psychiatry, 2015 May 1;77(9):823-32.

Malkova et al., 2012 Maternal immune activation yields offspring displaying mouse versions of the three core symptoms of autism, Brain Behaviorand Immunity, 2012 May ; 26(4): 607–616.

Marques et al., 2014 Autism Spectrum Disorder Secondary to Enterovirus Encephalitis, Journal of Child Neurology, 2014, Vol. 29(5) 708-714.

Mestas et al., 2004 Of Mice and Not Men: Differences between Mouse and Human Immunology, Journal of Immunology, 0022-1767:2731-2738.

Meyer et al., 2006 The Time of Prenatal Immune Challenge Determines the Specificity of Inflammation-Mediated Brain and Behavioral Pathology, The Journal of Neuroscience, 26(18):4752–4762.

Meyer et al., 2007 The neurodevelopmental impact of prenatal infections at different times of pregnancy: the earlier the worse?, Neuroscientist, Jun;13(3):241-56.

Meyer et al., 2009 In-vivo rodent models for the experimental investigation of prenatal immune activation effects in neurodevelopmental brain disorders, Neuroscience and Biobehavioral Reviews, 33 (2009) 1061–1079.

Meyer 2014, Prenatal Poly(I:C) Exposure and Other Developmental Immune Activation Models in Rodent Systems, Biological Psychiatry, 75:307-315.

Mitkus et al., 2011 Updated aluminum pharmacokinetics following infant exposures through diet and vaccination, Vaccine 29 (2011) 9538–9543.

Offit et al., 2003 Addressing Parents' Concerns: Do Vaccines Contain Harmful Preservatives, Adjuvants, Additives, or Residuals? Pediatrics, 112(6): 1394-1401.

Oskvig et al., 2012 Maternal immune activation by LPS selectively alters specific gene expression profiles of interneuron migration and oxidative stress in the fetus without triggering a fetal immune response, Brain Behavior and Immunity, 2012 May ; 26(4): 623–634.

Pang et al., 2016 Exploiting macrophages as targeted carrier to guide nanoparticles into glioma, Oncotarget 7(24):37081.

Parker-Athill and Tan, 2010 Maternal Immune Activation and Autism Spectrum Disorder: Interleukin-6 Signaling as a Key Mechanistic Pathway, NeuroSignals, 2010;18:113–128.

Petrik et al., 2007 Aluminum Adjuvant Linked to Gulf War Illness Induces Motor Neuron Death in Mice, NeuroMolecular Medicine, Vol. 9, 83-100.

Pineda et al., 2013 Maternal immune activation promotes hippocampal kindling epileptogenesis in mice, Annals of Neurology, 2013 July ; 74(1): 11–19.

Podila et al., 2013 Toxicity of Engineered Nanomaterials: A Physicochemical Perspective, Journal of Biochemical and Molecular Toxicology, 2013 January ; 27(1): 50–55.

Robertson et al., 2016 Reduced GABAergic Action in the Autistic Brain, Current Biology, 26, 1-6.

Saad et al., 2016 Vitamin D status in autism spectrum disorders and the efficacy of vitamin D supplementation in autistic children, Nutritional Neuroscience, 19 (8) 346-351.

Semple et al., 2013 Brain development in rodents and humans: Identifying benchmarks of maturation and vulnerability to injury across species, Progress in Neurobiology, Jul-Aug;106-107:1-16.

Sethi et al., 2008 Aluminium-induced electrophysiological, biochemical and cognitive modifications in the hippocampus of aging rats, Neurotoxicology 29, 1069-1079.

Sethi et al., 2009 Curcumin attenuates aluminium-induced functional neurotoxicity in rats, Pharmacology, Biochemisatry, and Behavior 93:31-39.

Shen et al., 2016 Postnatal activation of TLR4 in astrocytes promotes excitatory synaptogenesis in hippocampal neurons, Journal of Cell Biology, 215(5):719-734.

Sharifi et al., 2012 Toxicity of Nanomaterials, Chemical Society Reviews, 2012 Mar 21; 41(6): 2323–2343.

Shaw and Petrik, 2009 Aluminum hydroxide injections lead to motor deficits and motor neuron degeneration, Journal of Inorganic Biochemistry 103 (11).

Shaw and Tomljenovic, 2013 Administration of aluminium to neonatal mice in vaccinerelevant amounts is associated with adverse long term neurological outcomes, Journal of Inorganic Biochemistry, 128 (2013) 237–244.

Shi et al., 2009 Activation of the Maternal Immune System Alters Cerebellar Development in the Offspring, Brain, Behavior, and Immunity, January, 23(1): 116–123.

Smith et al., 2007 Maternal Immune Activation Alters Fetal Brain Development through Interleukin-6, Journal of Neuroscience, 2007 October 3; 27(40).

Smith et al., 2012, Maternal Immune Activation Increases Neonatal Mouse Cortex Thickness and Cell Density, Journal of Neuroimmune Pharmacology, 7(3):529-532.

Stiles et al., 2010 The Basics of Brain Development, Neuropsychology Reviews (2010) 20:327–348.

Suzuki et al., 2011 Plasma Cytokine Profiles in Subjects with High-Functioning Autism Spectrum Disorders, PloS ONE 6(5).

Suzuki et al., 2013 Microglial Activation in Young Adults With Autism Spectrum Disorder, JAMA Psychiatry 70(1): 49-58.

Takano 2015 Role of Microglia in Autism: Recent Advances, Developmental Neuroscience, 37:195-202.

Tau and Peterson, 2010 Normal Development of Brain Circuits, Neuropsychopharmacology, (2010) 35:147–168.

Taylor et al., 2014 Vaccines are not associated with autism: An evidence-based metaanalysis of case-control and cohort studies, Vaccine, 32:3623-3629.

Tomljenovic and Shaw, 2011 Do aluminum vaccine adjuvants contribute to the rising prevalence of autism? Journal of Inorganic Biochemistry 105.

Tsilioni et al., 2015 Children with autism spectrum disorders, who improved with a luteolin-containing dietary formulation, show reduced serum levels of TNF and IL-6, Translational Psychiatry, 5, 647.

Vargas et al., 2005 Neuroglial Activation and Neuroinflammation in the Brain of Patients with Autism, Annals of Neurology, 2005;57:67–81.

Viezeliene et al., 2013 Selective induction of IL-6 by aluminum-induced oxidative stress can be prevented by selenium, Journal of Trace Elements in Medicine and Biology, 27:226-229.

Wei et al., 2011 IL-6 is increased in the cerebellum of autistic brain and alters neural cell adhesion, migration and synaptic formation, Journal of Neuroinflammation 2011, 8:52.

Wei et al., 2012 (a) Brain IL-6 elevation causes neuronal circuitry imbalances and mediates autism-like behaviors, Biochimica et Biophysica Acta, 1822 (2012) 831–842.

Wei et al. 2012 (b) Alteration of brain volume in IL-6 overexpressing mice related to autism, International Journal of Developmental Neuroscience, 30:554-559.

Wei et al., 2013 Brain IL-6 and autism, Neuroscience 252 (2013): 320–325.

Wei et al., 2016 Inhibition of IL-6 trans-signaling in the brain increases sociability in the BTBR mouse model of autism, Biochimica et Biophysica Acta, 1862(10):1918-1925.

Weir et al., 2015 Preliminary evidence of neuropathology in nonhuman primates prenatally exposed to maternal immune activation, Brain, Behavior, and Immunity, 48,139–146.

Williams et al., 2006 The Profile of Memory Function in Children With Autism, Neuropsychology, 20(1): 21-29.

Wobke et al., 2014 Vitamin D in inflammatory diseases, Frontiers in Physiology, 5: 244.

Zerbo et al., 2014 Neonatal cytokines and chemokines and risk of Autism Spectrum Disorder: the Early Markers for Autism (EMA) study: a case-control study, Journal of Neuroinflammation, 11:113.

Zerbo et al., 2017 Association Between Influenza Infection and Vaccination During Pregnancy and Risk of Autism Spectrum Disorder, JAMA Pediatrics, 171(1).